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Applicants: Kang, C.-Y. *et al.*

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By: 
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**DECLARATION UNDER 37 C.F.R. § 1.132
BY CHIL-YONG KANG**

I, Chil-Yong Kang, a citizen of Canada, hereby declare as follows:

1. I am presently a Professor of Virology at the University of Western Ontario, Canada.
2. I have co-authored or authored over 130 publications and am a co-inventor of 4 issued U.S. patents in virology and vaccine development.
3. I hold a Ph.D. in virology from McMaster University in Hamilton, Ontario, Canada.
4. I am a co-inventor of U.S. Patent Application No. 09/762,294, filed April 2, 2002.

5. I have read the present Office Action and understand that the Examiner has rejected claims 1, 6, 7, and 31-36 stating that “it would have been *prima facie* obvious to one having ordinary skill in the art at the time the invention was made to prepare recombinant HIV-1 viruses with modified signal sequences, as taught by Li, et al. (1994), and to further include a *nef*-deletion in the construct, as provided by Daniel, et al. (1992), since this would provide a recombinant virus that is replication-impaired and expressed to high quantities.”

6. The recombinant HIV-1 virus that is the subject of and claimed in U.S. Patent Application No. 09/762,294 is avirulent, essentially non-cytolytic *and capable of highly efficient replication*. It can be also be expressed to high quantities. These properties are disclosed throughout the specification.

7. Attached as Exhibit A are the results of additional experiments performed to measure the viral production, infectivity and cytopathic effect of the NL4-3^{WT}, NL4-3^{nef-}, NL4-3^{SSR} and NL4-3^{SSR/nef-} viruses produced as described in Examples 5 and 6 of the specification of U.S. Patent Application No. 09/762,294. The data demonstrates that the signal sequence replaced (NL4-3^{SSR}) *and* combination *nef*-deleted/signal sequence replaced (NL4-3^{SSR/nef-}) viruses are able to be produced in high quantities, exhibit substantially reduced infectivity, and exhibit very little sign of cytopathic effect *despite active HIV replication*.

8. I am a co-author of Li, et al. (1994). This work shows only that the gp120 gene can be expressed in a large quantities in insect cells and secreted efficiently if the wildtype signal sequence is replaced with a honeybee mellitin signal sequence or IL-3 signal sequence. It does *not* teach a recombinant HIV-1 virus, let alone a recombinant HIV-1 virus that is avirulent, is essentially non-cytolytic and capable of highly efficient replication.

9. Daniel, et al. (1992) discloses a live attenuated SIV vaccine comprising a *nef*-deleted SIV. Daniel discloses that the *nef*-deleted SIV replicates poorly (see, for example, the Abstract). An accompanying paper to the Daniel reference, *Science* (1992) Dec 18;258(5090):1880-1, attached as Exhibit B, underscores the point that the Daniel attenuated virus “doesn’t replicate much” (page 1881, middle column). Further, Daniel does not describe large quantity SIV

production by *nef* deletion. Thus, combining the teaching of Daniel with that of Li *would not* produce a recombinant HIV-1 virus that is avirulent, essentially non-cytolytic and *capable of highly efficient replication*. Indeed, the article attached as Exhibit B cautions that SIV is different from HIV and that Daniel's monkey data may not apply to humans (page 1881, first column, second full paragraph). Thus, assuming one would even be motivated to apply the SIV vaccine techniques of Daniel in the context of HIV-1 in the light of such teaching, combining the teaching of Daniel with the teaching of Li would lead one to expect that at most deleting *nef* in HIV-1 would lead to a virus that replicates poorly.

10. To my knowledge, no one has generated a recombinant HIV-1 that is *avirulent, essentially non-cytolytic and capable of highly efficient replication* by inserting and/or substituting its genomic sequence. We are the first to show that *nef*-deletion, *vpu*-deletion and glycoprotein signal sequence replacement in the HIV-1 genome produces a recombinant HIV-1 that is able to be expressed in large quantities and that has the unexpected properties of being avirulent, essentially non-cytolytic *and* capable of highly efficient replication.

11. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

November 16, 2005

Date

Chil-Yong Kang
Chil-Yong Kang, Ph.D.

Exhibit A

Measurements of Viral Production

A3.01 cells were initially seeded into 6-well plates at a density of 1×10^6 cells/well and transfected with 10 μ g of proviral DNA. At 3 days post transfection, and every 2 days following, cultures were harvested and cells split 1:2 into fresh media without the addition of supplemental, uninfected cells. Harvested culture supernatants were pooled at each timepoint shown and analyzed for the presence of p24 by ELISA as indicator of virus production. Cells infected with either the NL4-3^{WT} or NL4-3^{nef} virus showed maximum virus production at 13 days post transfection, however cells showed high levels of CPE and cell numbers declined rapidly with cultures being discontinued by 17 days post transfection. Cells infected with either the NL4-3^{SSR} or NL4-3^{nef/SSR} viruses however, showed minimal CPE and remained persistently infected up to 29 days post transfection, at which point cells did eventually succumb to virus-induced CPE and cultures were discontinued. NL4-3^{WT} or NL4-3^{nef} virus cultures produced a maximum of 1×10^2 μ g p24 while NL4-3^{SSR} or NL4-3^{nef/SSR} viruses produced over 1×10^6 μ g p24 in a single harvest. Results shown in **Figure 1**.

Measuring Infectivity

Following transfection of proviral DNA, cells were split every 2 days and samples of the culture supernatant collected and analyzed by p24 ELISA in order to monitor viral replication. To assess the infectivity of virus particles being produced, samples were further analyzed by MAGI assay at 8 days post transfection in both A3.01 and H9 cells, and the results standardized to represent the number of infectious viral particles present per ng of p24 protein. As shown above, the Env signal sequence replacement mutant (NL4-3^{SSR}) and combination *nef*-deleted/Env signal sequence replacement mutant (NL4-3^{nef/SSR}) both possess substantially reduced infectivity, with the replacement mutant being approximately 2-fold to 3-fold less infectious than wild-type virus (NL4-3^{WT}), and the combination mutant exhibiting as much as a 50-fold decrease in infectivity as compared to the wild-type. Results shown in **Figure 2**.

Induction of Cytopathic Effect

H9 cells were infected at a multiplicity of infection 3 with each of the viruses indicated. Infections were allowed to proceed with cultures being split 1:2 every 2 days. At 6 days post infection, cells were examined by light microscopy and cytopathic effect (CPE) was observed. See **Figure 3**. As shown, H9 cells infected with either the NL4-3^{WT} or NL4-3^{nef} virus (both of which contain the natural Env signal sequence) exhibited a rapid onset of CPE including cell death and formation of large syncytia (black arrows). In contrast, cells infected with either the NL4-3^{SSR} or NL4-3^{nef-SSR} viruses (which contain the mellitin signal sequence in place of the natural Env signal sequence) showed very little sign of CPE despite active HIV replication (as measured by HIV-1 p24 ELISA; not shown).

FIGURE 1

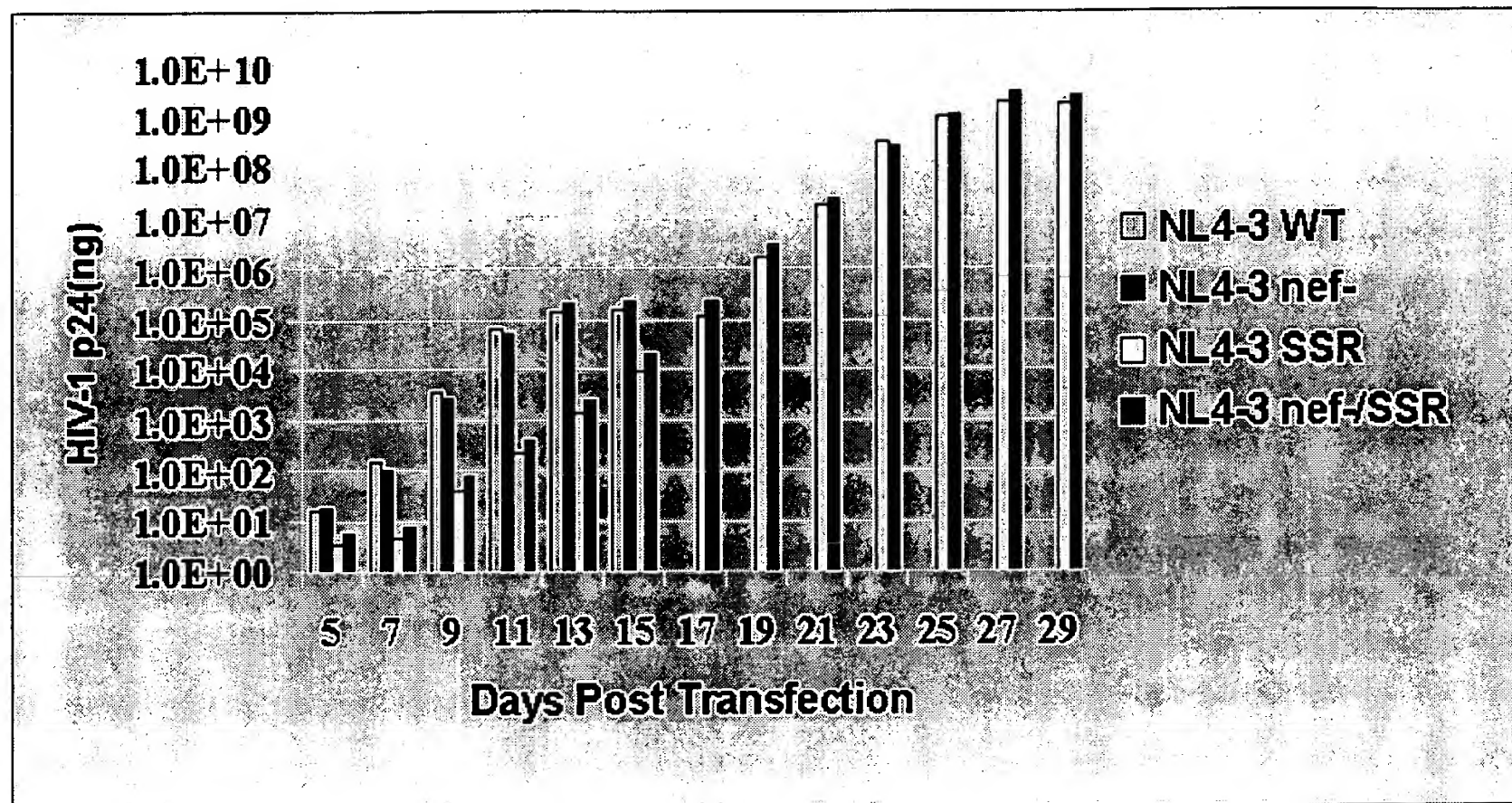
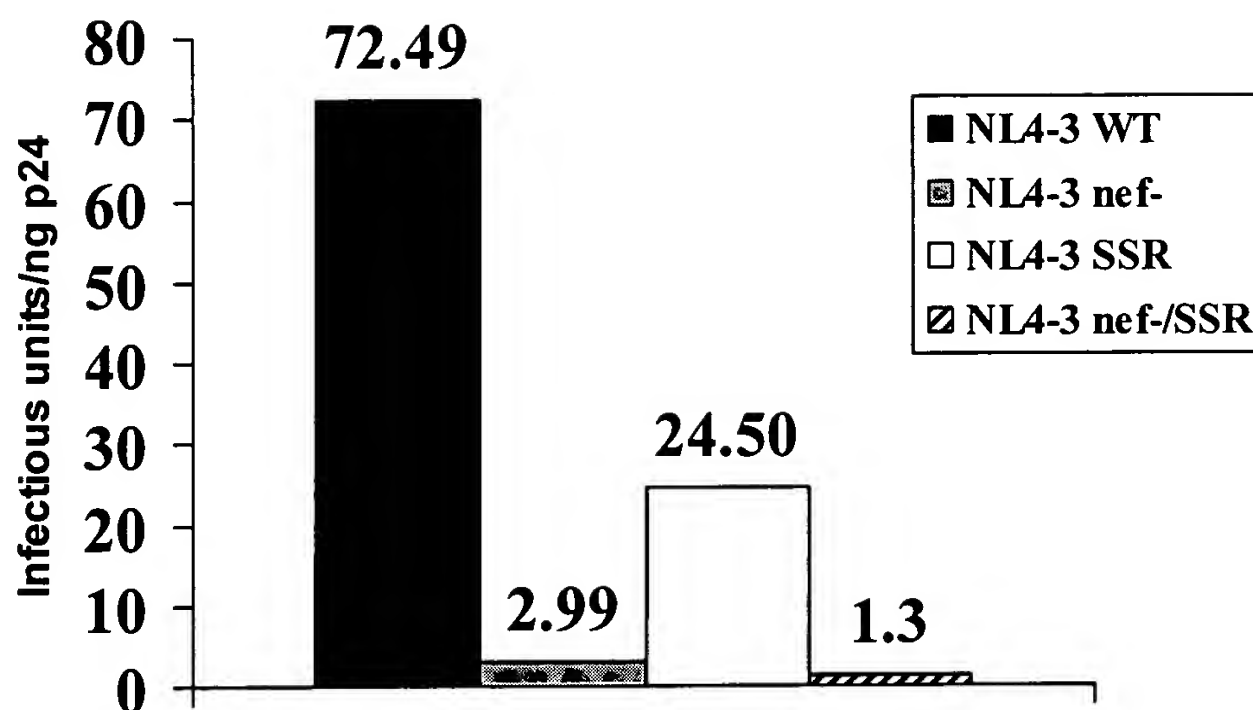


Figure 1 depicts a graph showing that prolonged cell survival leads to increased virus yield in A3.01 cells. Genetically modified HIV-1 clade B virus include NL4-3^{nef-}, which contains a targeted deletion of the *nef* gene, NL4-3^{SSR}, which has had the natural Env glycoprotein signal sequence replaced with the honeybee mellitin signal sequence, NL4-3^{nef-/SSR}, which contains a combination of both the *nef* deletion and signal sequence replacement mutations, and NL4-3^{WT}, which represents the parental, wild-type provirus.

FIGURE 2

A3.01 Cells



H9 Cells

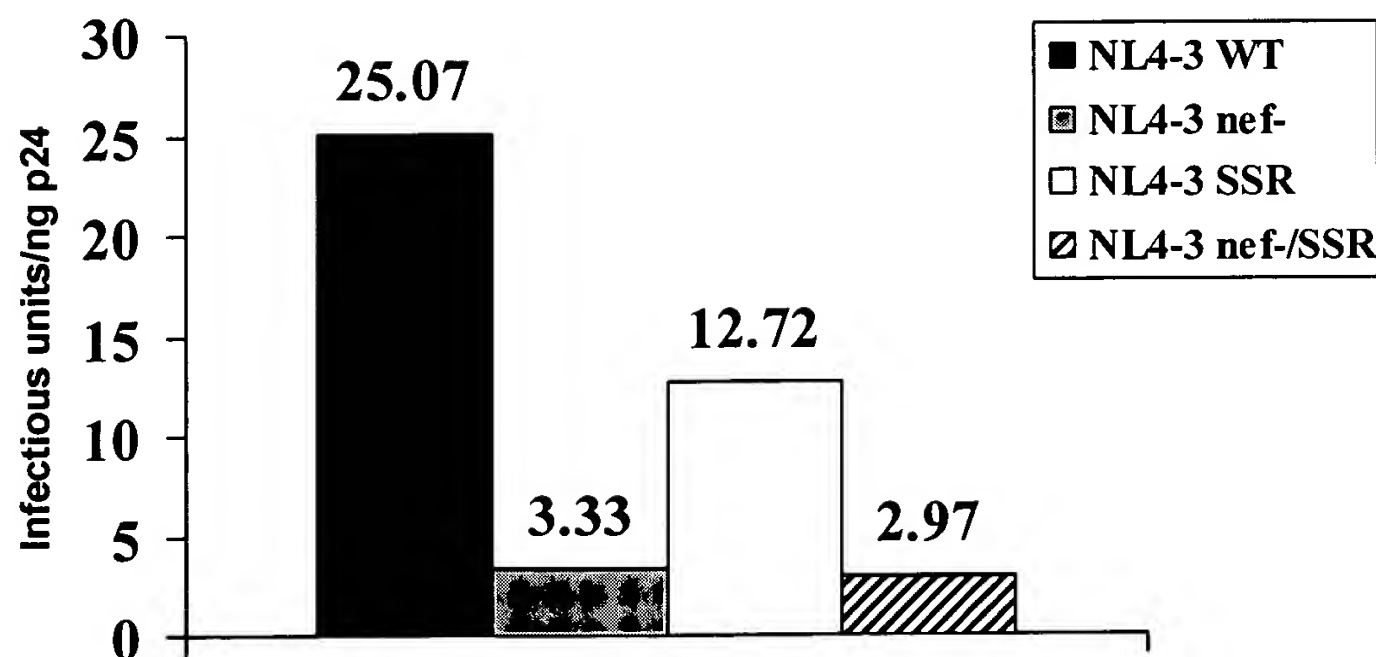


Figure 2 depicts the infectivity of HIV-1 NL4-3 mutants in A3.01 and H9 cells using the MAG1 assay. Genetically modified HIV-1 clade B virus include NL4-3^{*nef*⁻}, which contains a targeted deletion of the *nef* gene, NL4-3^{SSR}, which has had the natural Env glycoprotein signal sequence replaced with the honeybee mellitin signal sequence, NL4-3^{*nef*⁻/SSR}, which contains a combination of both the *nef* deletion and signal sequence replacement mutations, and NL4-3^{WT}, which represents the parental, wild-type provirus.

FIGURE 3

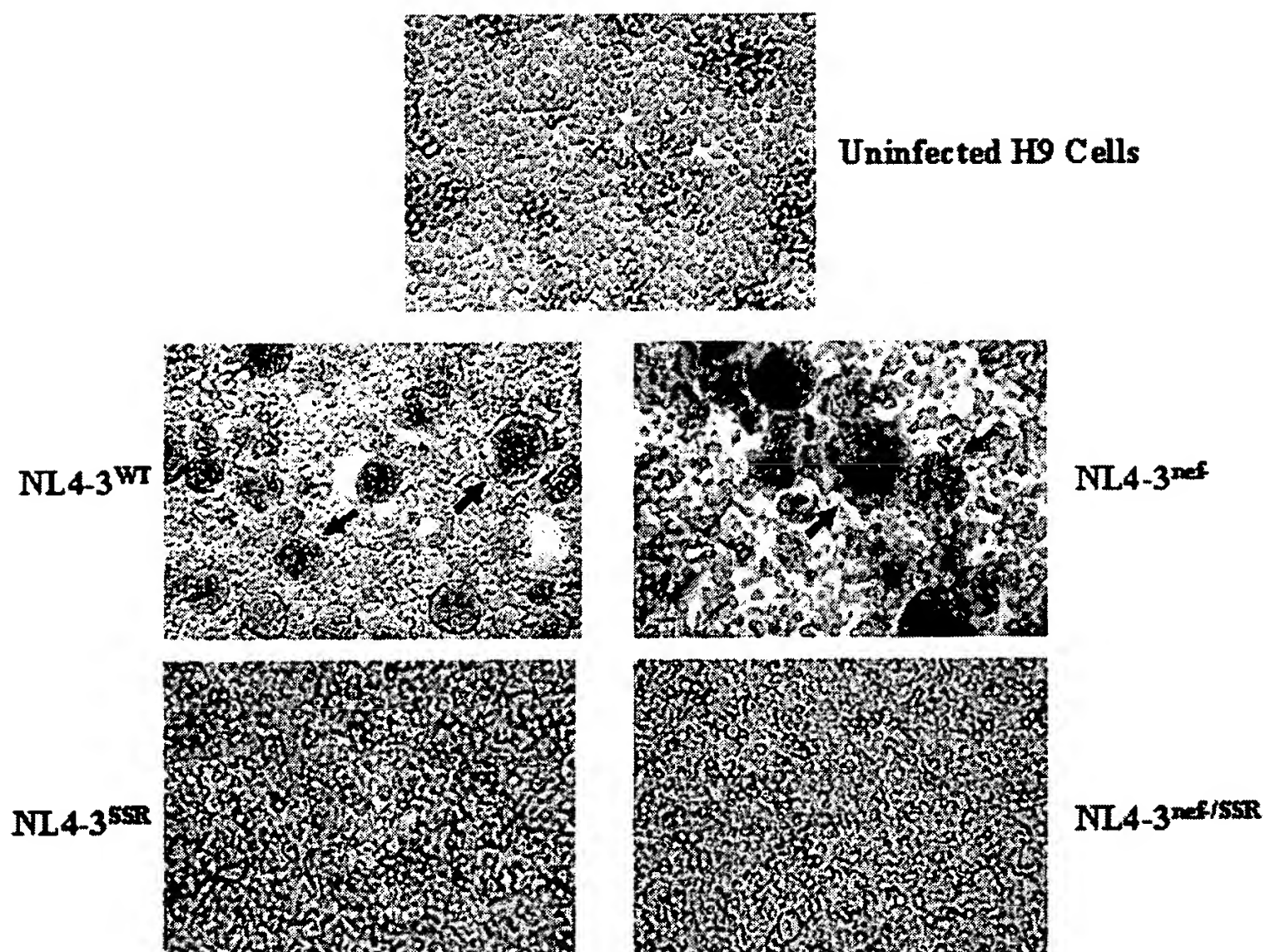


Figure 3 depicts the induction of cytopathic effect (syncytium formation) by HIV-1_{NL4-3} in H9 infected cells. Genetically modified HIV-1 clade B virus include NL4-3^{nef}, which contains a targeted deletion of the *nef* gene, NL4-3^{SSR}, which has had the natural Env glycoprotein signal sequence replaced with the honeybee mellitin signal sequence, NL4-3^{nef/SSR}, which contains a combination of both the *nef* deletion and signal sequence replacement mutations, and NL4-3^{WT}, which represents the parental, wild-type provirus.

RESEARCH NEWS

AIDS Vaccines: Is Older Better?

Researchers once wrote off vaccines based on live, weakened virus as far too dangerous, but recent data are causing some to rethink the old-fashioned approach

Scores of academic, government, and corporate labs around the world have entered the race to find an effective vaccine to prevent AIDS. But most are backing the same horse: genetically engineered pieces of the AIDS virus that they hope will be enough to trigger a protective immune response. In putting their bets on these engineered proteins, researchers have avoided the method used to develop human vaccines against most other viral diseases—a weakened version of the entire virus. AIDS vaccine developers recognize the power of attenuated virus vaccines, but they fear that even a weakened version of the stealthy and cunning HIV could lead to fatal infection.

A couple of recent developments have given researchers second thoughts, however, and some are now wondering whether they should hedge their bets. The first development is that the new-fangled, high-tech approach isn't working well: Human and animal experiments have yielded precious little data suggesting that vaccines made of HIV pieces will provide protection against the virus itself. The second is that an experiment involving attenuated virus has produced some startling results. On page 1938 of this issue of *Science*, Ronald Desrosiers and his colleagues at Harvard's New England Regional Primate Research Center report the longest-lasting, strongest protection yet achieved in any AIDS vaccine experiment—using precisely that old-fashioned method.

More than 2 years after vaccinating four rhesus monkeys once with a weakened form of SIV (a close simian relative of HIV), Desrosiers "challenged" them with a low-dose injection of full-strength virus—and all the monkeys resisted infection. What is more, the monkeys later resisted another challenge with a much higher dose of SIV. "This is a significant advance," says Anthony Fauci, director of the National Institute of Allergy and Infectious Diseases (NIAID). Alan Schultz, chief of the vaccine branch at NIAID's Division of AIDS, adds that "this is three orders of magnitude better than any protection we've seen." And Duke University's Dani Bolognesi calls the Desrosiers data "head

and shoulders above everything else."

Desrosiers' findings don't mean that the AIDS vaccine community is about to switch horses and make attenuated HIV vaccines. Plenty of safety concerns remain, and it could turn out that the live, attenuated approach does no more than provide cues for a safer vaccine based on engineered proteins. But in a crucial field, where the outlook has been

January, p. 456), although a handful of vaccines containing recombinant HIV proteins have shown promise in chimpanzee tests.

Desrosiers himself tried—and failed—with the recombinant protein approach. He also had only marginal success with the so-called whole, killed virus method, in which a genetically inactivated version of the entire virus is used. Aside from the fact that the results from whole, killed SIV vaccine experiments have been difficult to sort out (see sidebar), many researchers dismiss this approach, too, for safety reasons, noting that if any genetic material remains intact, the vaccine could cause AIDS.

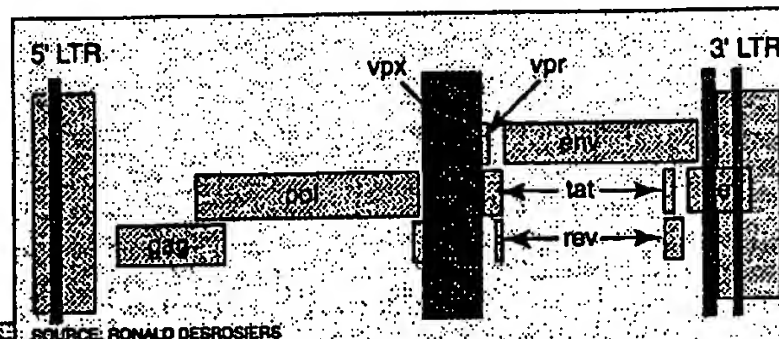
Desrosiers stumbled onto the live, attenuated approach while studying the mechanism by which the AIDS virus wreaks its damage. In that work, he

had constructed a clone of a highly virulent SIV strain that did not include a viral gene called *nef*. Three years ago, he gave six rhesus monkeys an injection of this *nef*-deleted SIV to see how the deletion affected the course of the disease and simultaneously gave another group unaltered SIV. While the monkeys that got the natural virus began dying of AIDS, the six who received the *nef*-deleted strain remained healthy.

Apparently, the *nef* deletion had somehow rendered the virus harmless. If that was the case, Desrosiers wondered, could he have inadvertently created a vaccine? To find out, he challenged four of the "vaccinated" monkeys and four control animals with a small dose of infectious SIV. Within 36 weeks, the four controls were dead or sick, whereas the four primed with the *nef*-mutant strain remained healthy. The strain of virus used for the challenge couldn't be detected in the monkeys' blood, even with ultrasensitive tests, indicating that they had managed to resist infection.

Desrosiers had shown that the inoculated animals could resist a relatively low dose of SIV, but could they fight off a serious challenge? The answer, so far, appears to be yes. Desrosiers injected the monkeys with a huge dose of infectious SIV, and after 18 weeks these animals show no evidence of infection.

Virologist Muthiah Daniel, first author of



Delete key. Deleting part of the genome of viruses that cause AIDS (deletions in red above) forms the basis of a vaccine strategy that has protected monkeys from infection.



gloomy, Desrosiers' results could provide hope, especially if his vaccine works in more rigorous trials.

Like all vaccines, one made from an attenuated live virus is designed to fool the immune system into reacting as if it were meeting

the actual, disease-causing organism. If the mock struggle works, the immune system will be primed to defeat the real enemy. What distinguishes the live viral vaccine from other approaches is that rather than simply presenting viral components, it actually causes infection. The virus replicates in the body, repeatedly providing a range of different proteins for the immune system to attack.

In contrast, the genetically engineered AIDS vaccines under commercial development typically consist of only a single HIV surface protein—most companies are focusing on a protein called gp120 or one called gp160. When mixed with enhancing chemicals, these single proteins can, in theory, prime the immune system without causing infection. To date, however, only one monkey experiment has had solid success with a recombinant protein SIV vaccine (*Science*, 24

the *Science* paper, cautions that "we're many years away before this can be used in humans," but Daniel says researchers should think seriously about this approach—because there isn't much choice. "The recombinant vaccines," he predicts, "are not going to work." Murray Gardner, a prominent monkey researcher at the University of California, Davis, who calls the new data "striking," is also pessimistic about vaccines made from one genetically engineered viral protein. "To think that a simple, recombinant protein can do the job is foolish," he says, because single proteins can't induce the full immune response needed to fight off a virus. Gardner has also been swayed by encouraging results obtained by his colleague Marta Marthas, using her own attenuated SIV vaccine.

The companies pursuing recombinant or chemically synthesized candidate vaccines (including Chiron, Genentech, MicroGeneSys, Immuno, Viral Technologies, and Pasteur Mérieux) obviously disagree with Gardner and Daniel. Yet the companies do stand to gain from the attenuated experiments, because they could provide crucial indications of how the immune system can prevent HIV infection. In that regard, as Duke's Bolognesi says, "so far, we've been flying without a compass."

Phillip Berman of Genentech hopes to apply lessons learned from the attenuated vaccine. But Berman cautions that SIV is different from HIV and that monkey data may not apply to humans. "SIV grows fast and intense in rhesus, where HIV is slow and smoldering in man," says Berman. And he also thinks there's strong evidence that a specific portion of gp120 called the V3 loop can trigger production of antibodies that can stop HIV. "The data's very good that you can protect chimpanzees from infection with antibodies to the V3 loop," he says.

Desrosiers says he hopes a recombinant vaccine or some other obviously safe method will prove effective. But he says: "Our backs are getting to the wall in a dramatic and dangerous situation. We need to be ready to accept some radical approaches." Before the live, attenuated approach could be seriously considered for use in humans, however, at least three safety concerns would have to be dealt with. First, the AIDS virus in an attenuated vaccine could revert to a virulent state. Second, even if it didn't cause AIDS, the vaccine might cause cancer. And a virus that initially looks safe might, decades later, turn out to cause disease.

Desrosiers has thought about all three concerns. Because the virus requires certain genetic elements to cause disease, he says, he is confident that if you remove enough of the viral genome, it would be practically impossible for an attenuated virus to revert to virulence; he is now testing mutants with up to five genetic deletions. As for a vaccine caus-

ing cancer, Robert Gallo of the National Cancer Institute argues that HIV is implicated in B cell lymphomas and Kaposi's sarcoma. But Desrosiers counters that the causal link isn't clear, and that, in any case, these cancers seem to be dependent on HIV replicating, and his attenuated virus doesn't replicate much.

Long-term safety concerns, however, are worrisome, concedes Desrosiers. "It will take 10 or 15 years of safety testing before we can be comfortable putting this into thousands of people." Which is why he thinks the process ought to begin soon. He's about to begin chimp trials of triple and quadruple deletion mutants. If those preparations prove safe and

give hints of efficacy, he believes tests should begin in a small number of human volunteers at high risk of HIV infection.

That's not a thrilling prospect, all involved concede. Yet some researchers believe it's time to rethink things. Patricia Fultz of the University of Alabama, Birmingham, who has tested AIDS vaccines in chimps and monkeys, says that 5 years ago she ruled out live vaccines. Now, she says, "we may be forced to use this as the only method that appears to have an impact." Which highlights the fact that, as in all AIDS research, the choices are between rocks and hard places.

—Jon Cohen

Explaining Puzzling Vaccine Results

A letter published in *Nature* last year by James Stott of England's National Institute for Biological Standards and Control didn't fill even one page—but it stood the AIDS vaccine world on its head. Stott described how his group had, as a control to a monkey vaccine experiment, injected four animals with human white blood cells. When the animals were "challenged" with virulent SIV (a monkey version of the AIDS virus) two failed to become infected, suggesting that something in the blood cells protected the monkeys. That worried researchers because human white blood cells are used to culture SIV and they have been used in many monkey vaccine trials—possibly skewing the results. In this issue of *Science* (page 1935), a team led by Larry Arthur of Program Resources Inc., a contractor with the National Cancer Institute, helps explain Stott's findings. But the new results also show just how wily the AIDS virus is in evading the immune system.



Studley. HIV studded with immune proteins.

The vaccine that has worked best in monkey experiments to date has been made from SIV whose genetic material is chemically or physically inactivated. Though researchers have serious safety concerns about this method (if any virus accidentally was not inactivated, the vaccine could cause AIDS), the failure-riddled AIDS vaccine field was heartened when this "whole, killed" virus approach began succeeding. Then Stott's letter threw into question the validity of every experiment involving whole, killed SIV vaccines. Were the immune responses stimulated by viral proteins in the vaccine or by an artifact?

Stott and his co-workers quickly came up with a theory. Like all retroviruses, SIV grows in cells and buds through their membranes, picking up cellular proteins in the process. Stott's study suggested that antibodies made to these cellular proteins—not to the SIV proteins—were the key to protection. Now Arthur spells out just what those cellular proteins are and how they could have provided the protection.

Arthur took purified HIV and SIV and separated out the cellular proteins, which include immune-system molecules called $\beta 2$ microglobulin ($\beta 2m$) and HLA DR. Strikingly, they found more $\beta 2m$ and HLA DR than HIV's own surface protein, gp120. Arthur also showed that concoctions of concentrated antibodies to $\beta 2m$ and HLA DR can block HIV and SIV infection in the test tube, supporting Stott's hypothesis. Arthur now intends to test an SIV vaccine made from cellular proteins alone.

Michael Murphey-Corb of the Tulane Regional Primate Research Center says that while cellular proteins explain some of the protection seen with whole, killed SIV vaccines, researchers should not write off the approach. "This whole inactivated vaccine worked too well not to be real," says Murphey-Corb, who is investigating other explanations for the Stott experiment.

—J.C.

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